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Abstract

A very significant challenge in Alzheimer's disease today is the discovery of suitable technologies for detection of the disease that are easy to use, cost effective and non-invasive. In this study we demonstrate that saliva biomarkers are a suitable option for detection of the disease. Accordingly, we collected saliva samples by three different methods from age and gender matched Alzheimer's patients and normal healthy subjects. Salivary A β -40, A β -42, IGF-I, IGF-II, alpha amylase, IL-1 β , and TNF-alpha levels were analyzed. Of the three methods, passive drooling was found to be the best saliva collection method for analysis of biomarkers of Alzheimer's disease. There were significant differences in the salivary biomarkers evaluated between patients and controls. Additional results from our studies confirm that A β -40, A β -42, IGF-I, IGF-II, alpha amylase, IL-1 β , and TNF-alpha are appropriate diagnostic biomarkers for Alzheimer's disease.

Key words: Saliva, Alzheimer's disease, biomarkers

Introduction

Alzheimer's disease [AD] is a lethal neurodegenerative disorder that presently affects approximately 10.6 million people in the USA and Europe, with predictable estimates reaching epidemic proportions of nearly 15.4 million afflicted by the year 2030 (1). AD leads to a decrease in cognitive function, loss of memory and other effects. It has been reported that more than US\$183 billion was spent on AD patients in 2011 in the USA alone new estimates that project cost increases to as high as US\$1 trillion by the year 2050 (1). For this reason there is an urgent need for early detection methodologies and effective treatment regimens. Confirmation of AD is only currently possible by postmortem analysis of brain specimens of dementia-afflicted subjects. The clinical detection of AD is based on a clinical examination which includes a battery of laboratory tests, functional neuro-imaging

tools such as functional MRI, PET etc, and neuropsychological evaluation by a range of different methods. Clinical symptoms only appear after the onset of disease (2-4). Certain β -amyloids (A β) such as A β 10, A β 12 etc., hyperphosphorylated tau protein, α -synuclein, ubiquitin, apolipoprotein E, alpha antichymotrypsin and others have been shown to be pathological indicators of AD (2-6).

A β 40, A β 42, apoE-2, apoE-3, apoE-4, interleukin-6, transforming growth factor β 1, monocyte chemo attractant protein-1, interferon α , interleukins-2, -3, heparin binding growth-associated molecule, macrophage inflammatory protein-1 β , interleukin-8 receptor B, nitric oxide synthase, macrophage-colony stimulating factor, fibroblast growth factor-9, interferon γ -inducible chemokine IP-10, vascular endothelial growth factor and others have also been proposed as biomarkers for AD based upon cerebrospinal fluid analysis (2-9). Different plasma and serum based biomarkers such as A β 40, A β 42, IgG, α -synuclein, phenylalanine, ITIH4, Gpx3, CDK5, TNF- α , total A β and others have also been observed to exhibit positive diagnostic properties (2,5, 10-13). Although there has been progress and advancements in the field of biomarkers for AD as mentioned above, and these have resulted in complete databases with protocols and published methods, AD biomarker research remains a relatively undeveloped area. Presently, there are no valid non-invasive biomarkers identified in patient samples that may be used with high sensitivity and specificity to diagnose AD. All of the above mentioned biomarker studies rely on invasive specimens, are expensive, require special training and can lead to possible infection. To overcome these challenges, the healthcare system requires a technology which is inexpensive, non-invasive, cost effective and easy to use. Recent advances include reports that saliva based technologies can meet these market requirements.

Very few studies are available on salivary biomarkers for AD and only a few suitable salivary biomarkers have been characterized up until now that have been validated for AD (14). In an important study five salivary biomarkers have been proposed for the diagnosis of AD (15), so we undertook further

research necessary to study these specific salivary biomarkers for AD in addition to the effect of different saliva collection methods on the recovery of these biomarkers.

Materials and methods

Fifteen (15) Alzheimer's disease (AD) patients and 10 non-demented controls without neurological disease were selected for this study. Informed consent was taken from each subject. Ethical permission was taken according to the Helsinki guidelines. All AD patients were diagnosed by using established literature methods (16-18). The matched age and gender control group consisted of family members of the AD patients who were selected and given clinical, cognitive and function examinations; however in this control group no neuro-imaging analyses were performed. Unstimulated saliva samples were taken from subjects in each group using simple "passive" drooling and also by two stimulated methods using commercially available saliva collection devices (Salivette® polyester roll device and the Salivette® cotton roll device, Sarstedt). The salivary biomarkers IL-1 β ; alpha amylase, A β -40, A β -42, IGF-I, IGF-II and TNF-alpha were quantified using Luminex USA, Salimetrics USA, Biosource International, Invitrogen, Van Wyk and Underwood antibody kits, respectively. Each of these biomarkers were evaluated as described in our previous work (15). Data were analyzed with SPSS version 11.0. The Spearman rank correlation method was used for correlation analyses.

Results

Salivary A β -40, A β -42, IGF-I and IGF-II were not detected using the Salivette® cotton roll based device in either of the patient groups. Levels of alpha amylase, IL-1beta and TNF-alpha levels were significantly lower in samples collected using the Salivette® cotton roll collection device in comparison to samples collected using the Salivette® polyester roll collection device. In each case the recoveries of the various biomarkers are lower than using the passive drooling technique in both groups (Table -1). Levels of salivary A β -40, A β -42, IGF-I, IGF-II, alpha amylase, IL-1 β , and TNF-alpha were all found to be significantly different in AD patients in comparison to normal, healthy controls (Table-1).

Discussions

Cotton based saliva collection methods have been confirmed to affect the levels of detectable biomarkers in saliva, such as A β -40, A β -42, IGF-I and IGF-II rendering these biomarkers undetectable in saliva. Further in this study we show that cotton based saliva collection leads to a decrease in levels of alpha amylase, IL-1 β and TNF-alpha, supporting earlier evidence from previous studies (19, 20). We postulate that the "cotton interference effect" may be due to the formation of a bond between certain salivary biomarkers and cotton fiber used to collect the specimens. We found significant differences in salivary A β -40, A β -42, IGF-I, IGF-II, alpha amylase, IL-1 β , and TNF-alpha levels in AD patients compared to normal healthy controls, supporting amyloid, inflammation and the infectious concept of AD pathology (2-13). Furthermore, IL-1 β , TNF-alpha and other biomarkers were not significantly elevated in HIV and other inflammatory diseases. We believe these biomarkers are suitably disposed to be used for the early diagnosis of AD. The fact that salivary levels of the biomarkers A β -40, A β -42, IGF-I and IGF-II were not detectable using cotton based collection devices in both groups also supports a previously reported study (21).

Limitations in this study are firstly that clinical and laboratory investigations were not done to confirm the status of the normal healthy controls due to a lack in financial support. Also there is a noticeable and broad range of biomarker levels in the AD patients and controls, and we believe this is due to our improper selection of controls (possibly due to the inclusion of a few dementia patients in the control group). In addition, we were unable to carry out all available diagnostic methods in order to confirm the diagnosis of AD in the dementia patient group, which may have contributed to the wide SD [Standard Deviation] and the overlap between AD patients and normal healthy individuals. A further conclusion of this brief study is that passive drooling represents the most appropriate of the three methods of saliva collection tested for the detection of AD biomarkers as well as neurological biomarkers in oral fluid [saliva] specimens.

Saliva as a convenient bodily fluid has unique advantages over serum, blood and CSF due to its non-invasive properties, ease of handling, simplicity and minimal training requirements. In addition saliva sampling is highly cost effective for the screening of large population (22) and deserves to find greater application in the future.

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References

1. Alzheimer's Association. 2011 Alzheimer's disease facts and figure
www.alz.org/national/documents/Facts_Figures_2011.pdf
2. Schneider P, Hampel H, Buerger K. Biological marker candidates of Alzheimer's disease in blood, plasma, and serum. *CNS Neurosci Ther.* 2009 Winter;15(4):358-74.
3. Mattman A, Feldman H, Forster B, Li D, Szasz I, Beattie BL, Schulzer M. Regional HmPAO SPECT and CT measurements in the diagnosis of Alzheimer's disease. *Can J Neurol Sci.* 1997 ;24(1):22-8.
4. Cedazo-Minguez A, Winblad B. Biomarkers for Alzheimer's disease and other forms of dementia: clinical needs, limitations and future aspects. *Exp Gerontol.* 2010 ;45(1):5-14.
5. Henriksen K, O'Bryant SE, Hampel H, Trojanowski JQ, Montine TJ, Jeromin A, Blennow K, Lönneborg A, Wyss-Coray T, Soares H, Bazenet C, Sjögren M, Hu W, Lovestone S, Karsdal MA, Weiner MW; Blood-Based Biomarker Interest Group. The future of blood-based biomarkers for Alzheimer's disease. *Alzheimers Dement.* 2013 Jul 11. pii: S1552-5260(13)00045-9.
6. Schapira AH. Recent developments in biomarkers in Parkinson disease. *Curr Opin Neurol.* 2013 Aug;26(4):395-400.
7. Lista S, Garaci FG, Ewers M, Teipel S, Zetterberg H, Blennow K, Hampel H. CSF A β 1-42 combined with neuroimaging biomarkers in the early detection, diagnosis and prediction of Alzheimer's disease. *Alzheimers Dement.* 2013 Jul 11. pii: S1552-5260(13)00657-2.
8. Richard E, Schmand BA, Eikelenboom P, Van Gool WA; Alzheimer's Disease Neuroimaging Initiative. MRI and cerebrospinal fluid biomarkers for predicting progression to Alzheimer's disease in patients with mild cognitive impairment: a diagnostic accuracy study. *BMJ Open.* 2013 Jun 20;3(6). pii: e002541. doi: 10.1136/bmjopen-2012-002541.
9. Blennow K, Zetterberg H. The application of cerebrospinal fluid biomarkers in early diagnosis of Alzheimer disease.. *Med Clin North Am.* 2013 May;97(3):369-76.
10. Koehler NK, Stransky E, Shing M, Gaertner S, Meyer M, Schreitmüller B, Leyhe T, Laske C, Maetzler W, Kahle P, Celej MS, Jovin TM, Fallgatter AJ, Batra A, Buchkremer G, Schott K, Richartz-Salzbürger E. Altered Serum IgG Levels to α -Synuclein in Dementia with Lewy Bodies and Alzheimer's Disease. *PLoS One.* 2013 May 31;8(5):e64649. doi: 10.1371/journal.pone.0064649.
11. Tanaka H, Shimazawa M, Takata M, Kaneko H, Tsuruma K, Ikeda T, Warita H, Aoki M, Yamada M, Takahashi H, Hozumi I, Minatsu H, Inuzuka T, Hara H. ITIH4 and Gpx3 are potential biomarkers for amyotrophic lateral sclerosis. *J Neurol.* 2013 Jul;260(7):1782-97.
12. Czapski GA, Maruszak A, Styczyńska M, ?ekanowski C, Safranow K, Strosznajder JB. Association between plasma biomarkers, CDK5 polymorphism and the risk of Alzheimer's disease. *Acta Neurobiol Exp (Wars).* 2012;72(4):397-411.
13. Huang CW, Wang SJ, Wu SJ, Yang CC, Huang MW, Lin CH, Cheng IH. Potential blood biomarker for disease severity in the taiwanese population with Alzheimer's disease. *Am J Alzheimers Dis Other Dement.* 2013 Feb;28(1):75-83. doi: 10.1177/1533317512467674.
14. Shi M, Sui YT, Peskind ER, Li G, Hwang H, Devic I, Gingham C, Edgar JS, Pan C, Goodlett DR, Furay AR, Gonzalez-Cuyar LF, Zhang J. Salivary tau species are potential biomarkers of Alzheimer's disease. *J Alzheimers Dis.* 2011;27(2):299-305.
15. US 20120295281 A1. Specific salivary biomarkers for risk detection, early diagnosis, prognosis and monitoring of alzheimer's and parkinson's diseases.
16. McKhann G, Drachman D, Folstein M, Katzman R, Price D, Stadlan EM. Clinical Diagnosis of Alzheimer's Disease: Report of the NINCDS-ADRDA Working Group Under the Auspices of the Department of Health and Human Services Task Force on Alzheimer's Disease. *Neurology.* 1984; 34: 939-944.
17. American Psychiatric Association: DSM-IV: Diagnostic and Statistical Manual of Mental Disorders, Washington DC.
18. Folstein MF, Folstein SE, McHugh PR. Mini-mental State -A Practical Method for Grading the Cognitive State of Patients for the Clinician. *J Psychiatric Research.* 1975. 12: 189-198.
19. Hansen AM, Garde AH, Persson R. Measurement of salivary cortisol--effects of replacing polyester with cotton and switching antibody. *Scand J Clin Lab Invest.* 2008;68(8):826-9.
20. Shirtcliff EA, Granger DA, Schwartz E, Curran MJ. Use of salivary biomarkers in biobehavioral research: cotton-based sample collection methods can interfere with salivary immunoassay results.

Psychoneuroendocrinology. 2001 Feb;26(2):165-73.

21. Shi M, Sui YT, Peskind ER, Li G, Hwang H, Devic I, Ginghina C, Edgar JS, Pan C, Goodlett DR, Furay AR, Gonzalez-Cuyar LF, Zhang J. Salivary tau species are potential biomarkers of Alzheimer's disease. *J Alzheimers Dis.* 2011;27(2):299-305.

22. Castagnola M, Picciotti PM, Messana I, Fanali C, Fiorita A, Cabras T, Calò L, Pisano E, Passali GC, Iavarone F, Paludetti G, Scarano E. Potential applications of human saliva as diagnostic fluid. *Acta Otorhinolaryngol Ital.* 2011;31(6):347-57.

Illustrations

Illustration 1

Table 1: Salivary Biomarkers Levels in AD Patients and Normal Healthy Controls

Salivary Biomarkers	(Normal Healthy Patients)			AD Patients		
	Whole Saliva	Stimulated Saliva		Whole Saliva	Stimulated Saliva	
		Salivette® cotton roll	Salivette® polyester roll		Salivette® cotton roll	Salivette® polyester roll
Aβ-40 (pg/ml)	30.8(6.7)	-	21.7 (4.8)	22.1 (15.2)	-	17.2 (11.3)
TNF-α (pg/ml)	66.4(34.2)	44.3(22.4)	55.8 (31.3)	208.3(67.2)	176.3 (46.8)	189.2 (34.3)
IL-1-β (pg/ml)	44.6(32.2)	35.6(26.8)	34.8 (21.3)	156.3(34.7)	104.8 (12.3)	132.3 (41.2)
Aβ-42 (pg/ml)	3.8 (2.3)	-	3.2 (2.1)	8.5 (1.3)	-	7.3 (3.2)
IGF-I (ng/ml)	2.4 (1.3)	-	2.1 (1.3)	1.1 (0.8)	-	0.8 (0.5)
IGF-II (ng/dL)	3.5 (1.8)	-	3.1 (2.2)	2.3 (1.2)	-	2.1 (1.3)
Alpha-amylase(U/ml)	19.6 (8.9)	14.3 (8.4)	15.9 (8.2)	34.1 (11.5)	21.2 (11.3)	27.3 (12.2)